

The Utility of Circulating microRNAs as Biomarkers in Viral Infection-Associated Cancers

Laura A. Adamson-Small, Ph.D

Department of Pediatrics, University of Florida, Gainesville, FL

Email: ladamson@ufl.edu

Abstract

Since their discovery, the role of microRNAs (miRNAs) has been expanding. These small, non-coding RNAs have been associated with gene regulation by binding to 3' UTRs. Specific miRNA expression profiles are associated with cancer development and may serve as markers of malignancy. Many of these cancers are also associated with viral infections. As some viruses express their own miRNAs, research has begun to investigate if cellular or viral-encoded miRNAs could also serve as specific markers of these virus-associated cancers. Discovery of miRNAs in both tissues and serum have been reported in patients with virus positive tumors. These miRNAs may emerge as important, non-invasive diagnostic tools for detecting tumor development and progression and improving cancer treatment.

Keywords virus, cancer, miRNA, biomarker, serum

Introduction

It is estimated that approximately 20% of cancers worldwide are due to infectious agents (1). These pathogens contribute to multiple cancer types. The most common viruses associated with cancers include Epstein-Barr virus (EBV), hepatitis B virus (HBV), hepatitis C virus (HCV), human T-cell leukemia virus-1 (HTLV), human papillomaviruses (HPV), Kaposi's sarcoma herpes virus (KSHV), and human immunodeficiency virus, type 1 (HIV-1; 2-3). The problem with determining a causal relationship between many of these cancers and viral infection is that the tumors often appear years after the initial infection or the virus alone is not the causative agent of the cancer (4). Many of these infectious agents are prevalent at high percentages in the general population and thus require co-factors to contribute to cancer development. This makes use of the virus itself as a diagnostic tool for cancer development limited. Initial studies on virus-specific markers in cancers have focused on the

presence of viral transcripts and/ or antibodies, or changes in gene expression seen only in virally infected cells. The use of other, more specific biomarkers in response to viral infection may improve understanding of the relationship between these viruses and their associated cancers.

MicroRNAs (miRNAs) are small 19-24 nucleotide non-coding RNAs that bind to the 3' UTR of complementary transcripts and are important in transcriptional regulation. Analysis of miRNA expression levels has been described for use in cancer treatment, diagnosis, and characterization (5). These RNAs play an important role in regulating cellular gene expression. In cancers, these miRNAs often target oncogenes, immune regulatory genes, cell motility, or other important sequences for cellular transformation and survival. Alterations in miRNA levels have been reported in a multitude of viral infections (6). Furthermore, some viruses also encode their own miRNAs (7). Many studies have identified miRNA expression patterns that

are specific to viral infection or cancer development.

The possibility of using miRNAs as biomarkers was further expanded by the discovery of miRNAs in serum and plasma samples (8). Many of the cancers evaluated for tumor-specific miRNA expression patterns are also associated with virus infections. Detection of these virus-associated miRNAs may give information on pathways that can be targeted in cancer treatment, or act as a specific biomarker of cancer cells. Table 1 summarizes the viruses reviewed in this report, the cancers they are associated with, and miRNAs identified as potential serum biomarkers. This paper will further summarize studies investigating specific miRNAs as minimally invasive, specific biomarkers in the serum of patients with virus-related cancers.

Epstein-Barr virus, lymphoma, and nasopharyngeal carcinomas

Also referred to as human herpesvirus 4 (HHV-4), EBV is a common human DNA virus associated with multiple

cancers including Hodgkin's, non-Hodgkin's, and Burkitt's lymphoma as well as nasopharyngeal carcinoma (9). Many of these cancers occur in the conjunction with HIV co-infection. Infection is passed through the saliva, and the virus persists in latency for the remainder of a person's life. The role of EBV in these cancers is varied depending on cancer type, and tumor development is associated with expression of a multitude of viral transcripts, immune suppression, and/or co-infection with another virus (9). Because a large percentage of the world's population is infected with EBV, establishment of a biomarker distinguishing individuals who may develop EBV-specific cancers would improve early detection of these malignancies.

Using miRNA expression levels in normal versus cancerous cells, multiple viral and host miRNAs have been associated with EBV-infected tumors (10). To date, few studies have investigated the presence or correlation of these miRNAs in serum from patients with EBV-associated cancers. Wong et al., detected six EBV encoded

Table 1: Virus associated cancers and reported miRNA serum markers.

Virus	Associated Cancer	miRNA Serum Markers
EBV	Nasopharyngeal carcinoma	BART6-5p*, 6-3p*, 14*, 18-5p*, 19-3p*, 2-5p*
HBV	Hepatocellular carcinoma	2, 101, 122, 223, 375
HCV	Hepatocellular carcinoma	20a, 92a, 122, 618, 650
HTLV	Adult T-cell lymphoma	n/a
HPV	Cervical cancer	n/a
KSHV	Kaposi's sarcoma, Primary effusion lymphoma	17-92, 106b/25
HIV	Non-Hodgkin Lymphoma	223

*denotes viral-encoded miRNAs

miRNAs (BART6-5p, 6-3p, 14, 18-5p, 19-3p, and 2-5p) in serum that was significantly associated with nasopharyngeal carcinoma (NPC) diagnosis (11). Additionally, there was a positive correlation between the miRNA copy numbers detected in the tumor cells and those found in the serum. This suggests that changes seen in miRNA expression patterns in EBV positive tumors may also be potential biomarkers in serum samples. Furthermore, if future studies demonstrate that these miRNAs are a contributing factor in tumor development, they could act as a target for cancer treatment.

Hepatitis B and C infection in liver cancers

HBV and HCV are two distinct viruses associated with chronic liver disease and cancer. HBV is a DNA virus that infects approximately two billion people worldwide, resulting in over 350 million chronic infections and 600,000 deaths (12). In contrast, the single-stranded, positive-sense RNA virus HCV affects a smaller but still substantial percentage of the world's population with a prevalence of approximately 2% worldwide or 123 million people (13). Both infections are transmitted through blood products. Chronic infection with either virus can result in liver cirrhosis, a common risk factor for hepatocellular carcinoma (HCC) development. Recent evidence has detected an increase in mortality in cirrhosis-associated HCC cases, especially in patients infected with HCV (14). To determine which patients may be at risk for viral cirrhosis-associated liver cancer, studies have been conducted to investigate biomarkers correlated to HBV or HCV infection.

Evaluation of miRNA expression patterns in patient serum has revealed

several specific markers that may be deregulated only in HCC patients infected with HBV or HCV. The most common miRNAs in HBV and HCV infected patients have been identified as miR-21 (15), miR-101 (16), miR-223 (15), miR-375 (17), and miR-20a, (18), miR-92a (18), miR-618 (19), miR-650 (19), respectively. miR-122 was identified as a potential biomarker in both HBV and HCV-related cancers. (20-22). Interestingly, miR-122 has also been shown to be important in HCV infection by facilitating viral replication and protecting the viral genome from degradation, suggesting multiple roles for these miRNAs in virus-cell interaction (23-24). In patients with HBV-related HCC, serum levels of miR-122 following surgery were significantly reduced, suggesting this marker may also have potential utility in measuring cancer progression and recurrence (20).

Knowledge of these virus associated miRNAs that are deregulated could contribute to disease detection and treatment. Detection of the miRNA in the serum itself may be a specific biomarker for early detection of the cancer. Furthermore, if this miRNA is deregulated in both the serum and tumor, it may indicate a potential pathway for target treated. miR-21, which was shown to be upregulated in HBV-positive patients with HCC has been shown to modulated the PTEN pathway (25). Expression of certain miRNAs may also be used to evaluate treatment efficacy. Ji et al., (2009) reported an increased response to interferon treatment but lower overall survival in HCC patients with low miR-26 levels (26). Whether the miRNAs associated with viral expression could be utilized the same way has yet to be determined.

Human T-cell leukemia virus and adult T-cell leukemia and lymphoma

HTLV is a member of the retrovirus family that is aptly named because of its association with adult T-cell leukemia (ATL) and lymphoma. This RNA virus is estimated to infect 15-20 million people worldwide (27). Multiple groups have examined miRNA expression in HTLV infected cells, but reports examining miRNA expression profiles in serum could not be found. Cellular miRNAs associated with HTLV infection *in vivo* include miR-31 (28), miR-125b (29) and 9 other miRNAs identified *ex vivo* in cells from ATL patients (30). Future studies may determine if these miRNAs could also serve as serum biomarkers of HTLV associated cancer.

Human papillomaviruses

HPV is a DNA virus found in nearly 100% of cervical cancer cases (31). This virus has also been associated with oral, esophageal, and genital cancers. There are over 150 different HPV genomes variants classified that vary in phenotype and clinical presentation (32). miRNA analysis has been investigated in multiple studies of cervical tissue samples infected with HPV compared to normal controls (33-36). These papers detected a wide range of miRNA expression profiles associated with HPV infection and cancer with little overlap in results. The two miRNAs that were most frequently associated were miR-34a and miR-125b (37). While none of these studies examined serum, a large number of potential miRNAs have a potential association with HPV infection and cancer development.

Kaposi's sarcoma herpes virus

KSHV, also called HHV-8, is a DNA virus in the herpes virus family. KSHV malignancies depend on several factors including co-infection with HIV and state of the host immune response (38). KSHV not

only regulates host miRNA expression but also encodes its own miRNAs. To date, 24 KSHV miRNAs have been reported (39). These viral miRNAs contribute to tumor development in virally infected cells.

In an elegant study analyzing miRNA expression associated with Kaposi's sarcoma and primary effusion lymphoma, two of the same miRNA clusters were identified as altered in both the cancer patients tumor and serum samples compared to normal controls (40). miRNAs in the 17-92 and 106b/25 clusters containing miRNAs 17, 18, 19a, 19b, 20a, 25, 92a, 93, 93#, and 106b were investigated. While the largest changes in miRNA expression were in tumor samples, increased expression of these clusters and miRNAs 17, 19a, 19b, and 92a were in patient plasma samples. Because of the large number of KSHV encoded miRNAs, as well as studies demonstrating their importance in tumor development, research involving KSHV associated cancer and miRNA expression is rapidly growing.

Human immunodeficiency virus and non-Hodgkin lymphoma

The HIV virus is an RNA virus of the *Retroviridae* family (41). Its primary cellular target is CD4+ T cells, macrophages, and dendritic cells. HIV infection is currently reported in approximately 35.3 million people worldwide (41). Patients with HIV infection have a 60-200 fold increased risk of developing non-Hodgkin lymphoma (NHL) and a 8-10 fold increased risk of developing Hodgkin lymphoma (42). Other types of lymphomas, as well as anal, cervical, hepatocellular, and lung cancers are also more prevalent in HIV-positive patients, especially in those co-infected with KSHV. Defining key markers of viral-induced oncogenesis is an important step in

early detection towards improving the outcome of these patients.

In a study examining patients with NHL, four miRNAs (miR-21, 122, 222, and 223) were identified as specific to virus infection, but only miR-222 demonstrated potential diagnostic utility as a serum marker of both HIV infection and NHL development (43). Upregulation of miR-21 was reported in circulating B-cells derived from NHL patients, although not in serum (44). This marker was also identified in the previously listed study of serum from HIV-positive NHL patients. While this method is only applicable to circulating cancer cells, it further demonstrates the potential utility of miRNA expression as a marker of virally related cancer progression.

Potential use of virus-associated miRNAs in cancer management

The discovery of miRNAs in serum has revealed a new potential route for evaluating biomarkers in cancers and other diseases (8). These biomarkers could potentially play a role in early detection of cancers, for monitoring cancer progression, or evaluation of effective treatments. Circulating miRNAs have potential clinical

application due to the ease of testing serum samples (45). The largest virus-miRNA associations in serum studied to date involve miRNA detection in serum from HBV and HCV-associated cancers. These studies have reported miRNAs such as miR-122 which have altered expression in both primary tumors and in the serum in cancerous compared to normal controls and in relation to virus infection. Utility of miR-122 as both a biomarker and a therapeutic target in HCV-mediated liver cancer has been discussed (46). Identification of miRNA expression profiles in tumor tissues have multitudes of applications including understanding tumor development, improving treatment, and as reviewed in Figure 1, providing a potential tool for early cancer detection. Forthcoming studies will determine if miRNA serum profiles mimic those seen in cancerous tissue.

Multiple groups have investigated the use of serum miRNAs as biomarkers of cancer, metastases, Parkinson's disease, virus infection, heart disease, and some inflammatory conditions (47-49). Both serum and plasma could provide adequate samples for miRNA measurements, as they possess similar miRNA levels (50). It is

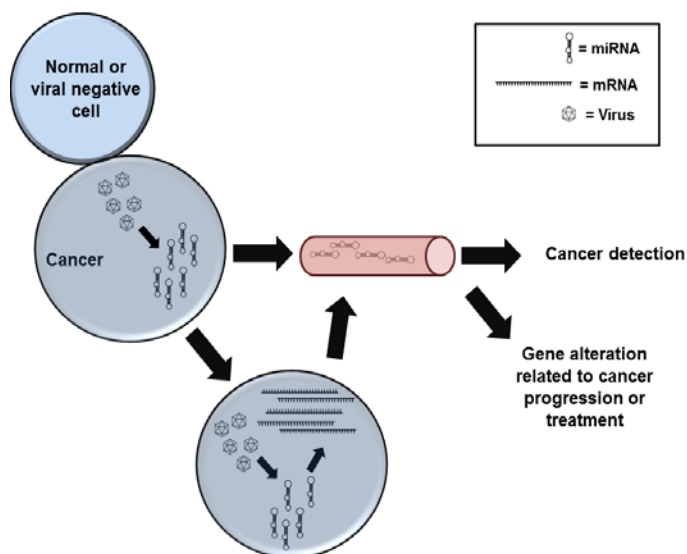


Figure 1: Summary of potential use of miRNAs in cancer management. Virus infection of cancer cells can alter miRNA expression. Changes in the same corresponding miRNA levels in serum samples could be used in early cancer detection and diagnosis. Changes in miRNA expression within cancer cells following viral infection may also alter cellular gene expression. If these genes impact cancer progression or cellular response to treatment, miRNAs in the serum may also be utilized in predicating cancer progression or response to treatment.

possible the circulating miRNAs detected in these studies are produced by cancer cells, as similar miRNA expression patterns have been reported in serum and tumors in both the EBV and KSHV studies reviewed here (11, 40). If further studies prove this to be true, many markers already identified as cancer-cell specific may be utilized as potential serum biomarkers.

Specific diagnostic tools are needed in virus related cancers. Many of the viruses reviewed here are prevalent at high frequencies in the human population, but only contribute to cancer under specific conditions. Improved diagnostic markers that can differentiate those susceptible to developing cancer following infection would have important clinical applications. Current virus diagnostic tools measure viral nucleic acids or proteins, or detect virus specific antibodies. While these can be useful in some cases, detection of the virus alone is not always able to distinguish individuals prone to cancer development. Additionally, if these circulating miRNAs are released from virus-positive cancer cells, they could be utilized as a marker of treatment efficacy and disease progression. These miRNAs may provide a novel, non-invasive method to distinguish between virally infected individuals and those susceptible to cancer development.

Acknowledgments

I would like to thank the Department of Pediatrics at the University of Florida for supporting my Postdoctoral research, and the reviewers for strengthening this manuscript.

References:

1. Parkin DM 2006 The global health burden of infection-associated cancers in

the year 2002. *International Journal of Cancer* **118**:3030-3044.

2. Bouvard V, Baan R, Straif K, Grosse Y, Secretan B, Ghissassi FE, Benbrahim-Tallaa L, Guha N, Freeman C, Galichet L, Cogliano V 2009 A review of human carcinogens—Part B: biological agents. *The lancet oncology* **10**:321-322.

3. Pagano JS, Blaser M, Buendia M-A, Damania B, Khalili K, Raab-Traub N, Roizman B 2004 Infectious agents and cancer: criteria for a causal relation. *Seminars in Cancer Biology* **14**:453-471.

4. Mesri Enrique A, Feitelson MA, Munger K 2014 Human Viral Oncogenesis: A Cancer Hallmarks Analysis. *Cell Host & Microbe* **15**:266-282.

5. Cho WCS 2010 MicroRNAs: Potential biomarkers for cancer diagnosis, prognosis and targets for therapy. *The International Journal of Biochemistry & Cell Biology* **42**:1273-1281.

6. Grassmann R, Jeang K-T 2008 The roles of microRNAs in mammalian virus infection. *Biochimica et Biophysica Acta (BBA) - Gene Regulatory Mechanisms* **1779**:706-711.

7. Pfeffer S, Zavolan M, Grässer FA, Chien M, Russo JJ, Ju J, John B, Enright AJ, Marks D, Sander C, Tuschl T 2004 Identification of Virus-Encoded MicroRNAs. *Science* **304**:734-736.

8. Chen X, Ba Y, Ma L, Cai X, Yin Y, Wang K, Guo J, Zhang Y, Chen J, Guo X, Li Q, Li X, Wang W, Zhang Y, Wang J, Jiang X, Xiang Y, Xu C, Zheng P, Zhang J, Li R, Zhang H, Shang X, Gong T, Ning G, Wang J, Zen K, Zhang J, Zhang C-Y 2008 Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res* **18**:997-1006.

9. Thompson MP, Kurzrock R 2004 Epstein-Barr Virus and Cancer. *Clinical Cancer Research* **10**:803-821.
10. Lopes LF, Ruiz Miyazawa KW, de Almeida ERD, Serafim KGG, de Almeida Gualtieri K, Costa IC, Felipe I, Pavanelli WR, Watanabe MAE 2013 Epstein-Barr Virus (EBV) MicroRNAs: Involvement in Cancer Pathogenesis and Immunopathology. *International Reviews of Immunology* **32**:271-281.
11. Wong AMG, Kong KL, Tsang JWH, Kwong DLW, Guan X-Y 2012 Profiling of Epstein-Barr virus-encoded microRNAs in nasopharyngeal carcinoma reveals potential biomarkers and oncomirs. *Cancer* **118**:698-710.
12. Shepard CW, Simard EP, Finelli L, Fiore AE, Bell BP 2006 Hepatitis B Virus Infection: Epidemiology and Vaccination. *Epidemiologic Reviews* **28**:112-125.
13. Shepard CW, Finelli L, Alter MJ 2005 Global epidemiology of hepatitis C virus infection. *The Lancet Infectious Diseases* **5**:558-567.
14. Fattovich G, Stroffolini T, Zagni I, Donato F 2004 Hepatocellular carcinoma in cirrhosis: Incidence and risk factors. *Gastroenterology* **127**:S35-S50.
15. Xu J, Wu C, Che X, Wang L, Yu D, Zhang T, Huang L, Li H, Tan W, Wang C, Lin D 2011 Circulating MicroRNAs, miR-21, miR-122, and miR-223, in patients with hepatocellular carcinoma or chronic hepatitis. *Molecular Carcinogenesis* **50**:136-142.
16. Fu Y, Wei X, Tang C, Li J, Liu R, Shen A, Wu Z 2013 Circulating microRNA-101 as a potential biomarker for hepatitis B virus-related hepatocellular carcinoma. *Oncology Letters* **6** : 1811-1815
17. Li L-M, Hu Z-B, Zhou Z-X, Chen X, Liu F-Y, Zhang J-F, Shen H-B, Zhang C-Y, Zen K 2010 Serum microRNA Profiles Serve as Novel Biomarkers for HBV Infection and Diagnosis of HBV-Positive Hepatocarcinoma. *Cancer research* **70**:9798-9807.
18. Shrivastava S, Petrone J, Steele R, Lauer GM, Di Bisceglie AM, Ray RB 2013 Up-regulation of circulating miR-20a is correlated with hepatitis C virus-mediated liver disease progression. *Hepatology* **58**:863-871.
19. Abdalla MA, Haj-Ahmad Y 2012 Promising candidate urinary microRNA biomarkers for the early detection of hepatocellular carcinoma among high-risk hepatitis C virus Egyptian patients. *Journal of Cancer* **3**:19.
20. Qi P, Cheng S-q, Wang H, Li N, Chen Y-f, Gao C-f 2011 Serum MicroRNAs as Biomarkers for Hepatocellular Carcinoma in Chinese Patients with Chronic Hepatitis B Virus Infection. *PLoS ONE* **6**:e28486.
21. Zhang Y, Jia Y, Zheng R, Guo Y, Wang Y, Guo H, Fei M, Sun S 2010 Plasma MicroRNA-122 as a Biomarker for Viral-, Alcohol-, and Chemical-Related Hepatic Diseases. *Clinical Chemistry* **56**:1830-1838.
22. Varnholt H, Drebber U, Schulze F, Wedemeyer I, Schirmacher P, Dienes H-P, Odenthal M 2008 MicroRNA gene expression profile of hepatitis C virus-associated hepatocellular carcinoma. *Hepatology* **47**:1223-1232.
23. Jopling CL, Yi M, Lancaster AM, Lemon SM, Sarnow P 2005 Modulation of Hepatitis C Virus RNA Abundance by a Liver-Specific MicroRNA. *Science* **309**:1577-1581.
24. Li Y, Masaki T, Yamane D, McGivern DR, Lemon SM 2013 Competing and noncompeting activities of miR-122 and the 5' exonuclease Xrn1 in regulation of hepatitis C virus replication. *Proceedings of*

the National Academy of Sciences **110**:1881-1886.

25. Meng F, Henson R, Wehbe-Janek H, Ghoshal K, Jacob ST, Patel T 2007 MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterology* **133**: 647-58

26. Ji J, Shi J, Budhu A, Yu Z, Forgues M, Roessler S, Ambs S, Chen Y, Meltzer PS, Croce CM, Qin LX, Man K, Lo CM, Lee J, Ng IO, Fan J, Tang ZY, Sun HC, Wang XW 2009 MicroRNA expression, survival, and response to interferon in liver cancer. *New England Journal of Medicine* **361**: 1437-47

27. Proietti FA, Carneiro-Proietti ABF, Catalan-Soares BC, Murphy EL 2005 Global epidemiology of HTLV-I infection and associated diseases. *Oncogene* **24**:6058-6068.

28. Yamagishi M, Nakano K, Miyake A, Yamochi T, Kagami Y, Tsutsumi A, Matsuda Y, Sato-Otsubo A, Muto S, Utsunomiya A, Yamaguchi K, Uchimarui K, Ogawa S, Watanabe T 2012 Polycomb-Mediated Loss of miR-31 Activates NIK-Dependent NF- κ B Pathway in Adult T Cell Leukemia and Other Cancers. *Cancer Cell* **21**:121-135.

29. Nicolette LDdF, Nicolette R, Haddad R, Azevedo R, Castro FAd, Tanaka Y, Takayanagui OM, Covas DT, Kashima S 2012 Upregulation of hsa-miR-125b in HTLV-1 asymptomatic carriers and HTLV-1-associated myelopathy/tropical spastic paraparesis patients. *Memórias do Instituto Oswaldo Cruz* **107**:824-827.

30. Bellon M, Lepelletier Y, Hermine O, Nicot C 2009 Deregulation of microRNA involved in hematopoiesis and the immune response in HTLV-I adult T-cell leukemia. *Blood* **113**:4914-4917.

31. Asiatic A, Ahmad ST, Mohammad SO, Zargar MA 2014 Review of the current

knowledge on the epidemiology, pathogenesis, and prevention of human papillomavirus infection. *European journal of cancer prevention : the official journal of the European Cancer Prevention Organisation (ECP)* **23**:206-224.

32. Bernard HU, Calleja-Macias IE, Dunn ST 2006 Genome variation of human papillomavirus types: phylogenetic and medical implications. *International journal of cancer Journal international du cancer* **118**:1071-1076.

33. Li Y, Liu J, Yuan C, Cui B, Zou X, Qiao Y 2010 High-risk human papillomavirus reduces the expression of microRNA-218 in women with cervical intraepithelial neoplasia. *The Journal of international medical research* **38**:1730-1736.

34. Li B, Hu Y, Ye F, Li Y, Lv W, Xie X 2010 Reduced miR-34a expression in normal cervical tissues and cervical lesions with high-risk human papillomavirus infection. *International journal of gynecological cancer : official journal of the International Gynecological Cancer Society* **20**:597-604.

35. Martinez I, Gardiner AS, Board KF, Monzon FA, Edwards RP, Khan SA 2008 Human papillomavirus type 16 reduces the expression of microRNA-218 in cervical carcinoma cells. *Oncogene* **27**:2575-2582.

36. Lajer CB, Garnaes E, Friis-Hansen L, Norrild B, Therkildsen MH, Glud M, Rossing M, Lajer H, Svane D, Skotte L, Specht L, Buchwald C, Nielsen FC 2012 The role of miRNAs in human papilloma virus (HPV)-associated cancers: bridging between HPV-related head and neck cancer and cervical cancer. *Br J Cancer* **106**:1526-1534.

37. Ribeiro J, Sousa H 2014 MicroRNAs as biomarkers of cervical cancer development: a literature review on miR-

125b and miR-34a. *Molecular biology reports* **41**:1525-1531.

38. Dittmer DP, Damania B 2013 Kaposi sarcoma associated herpesvirus pathogenesis (KSHV): an update. *Current Opinion in Virology* **3**:238-244.

39. Zhu Y, Haecker I, Yang Y, Gao S-J, Renne R 2013 γ -Herpesvirus-encoded miRNAs and their roles in viral biology and pathogenesis. *Current Opinion in Virology* **3**:266-275.

40. Chugh PE, Sin SH, Ozgur S, Henry DH, Menezes P, Griffith J, Eron JJ, Damania B, Dittmer DP 2013 Systemically circulating viral and tumor-derived microRNAs in KSHV-associated malignancies. *PLoS pathogens* **9**:e1003484.

41. Moss JA 2013 HIV/AIDS Review. *Radiologic Technology* **84**:247-267.

42. Carbone A, Vaccher E, Ghoghini A, Pantanowitz L, Abayomi A, de Paoli P, Franceschi S 2014 Diagnosis and management of lymphomas and other cancers in HIV-infected patients. *Nature reviews Clinical oncology* **11**:223-238.

43. Thapa DR, Hussain SK, Tran WC, D'Souza G, Bream JH, Achenback CJ, Ayyavoo V, Detels R, Martinez-Maza O 2014 Serum microRNAs in HIV-infected individuals as pre-diagnosis biomarkers for AIDS-related non-Hodgkin lymphomas (AIDS-NHL). *Journal of acquired immune deficiency syndromes (1999)*.

44. Thapa DR, Bhatia K, Bream JH, D'Souza G, Rinaldo CR, Wolinsky S, Detels R, Martinez-Maza O 2012 B-cell activation induced microRNA-21 is elevated in circulating B cells preceding the diagnosis of AIDS-related non-Hodgkin lymphomas. *AIDS (London, England)* **26**:1177-1180.

45. Cortez MA, Calin GA 2009 MicroRNA identification in plasma and

serum: a new tool to diagnose and monitor diseases. *Expert Opinion on Biological Therapy* **9**:703-711.

46. Pan QW, Henry SD, Scholte BJ, Tilanus HW, Janssen HL, van der Laan LJ 2007 New therapeutic opportunities for hepatitis C based on small RNA. *World journal of gastroenterology : WJG* **13**:4431-4436.

47. Gilad S, Meiri E, Yogev Y, Benjamin S, Lebanony D, Yerushalmi N, Benjamin H, Kushnir M, Cholakh H, Melamed N, Bentwich Z, Hod M, Goren Y, Chajut A 2008 Serum microRNAs are promising novel biomarkers. *PLoS One* **3**:e3148.

48. Chen X, Ba Y, Ma L, Cai X, Yin Y, Wang K, Guo J, Zhang Y, Chen J, Guo X, Li Q, Li X, Wang W, Zhang Y, Wang J, Jiang X, Xiang Y, Xu C, Zheng P, Zhang J, Li R, Zhang H, Shang X, Gong T, Ning G, Wang J, Zen K, Zhang J, Zhang CY 2008 Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res* **18**:997-1006.

49. Botta-Orfila T, Morato X, Compta Y, Lozano JJ, Falgas N, Valdeoriola F, Pont-Sunyer C, Vilas D, Mengual L, Fernandez M, Molinuevo JL, Antonell A, Marti MJ, Fernandez-Santiago R, Ezquerra M 2014 Identification of blood serum micro-RNAs associated with idiopathic and LRRK2 Parkinson's disease. *Journal of neuroscience research*.

50. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, Peterson A, Noteboom J, O'Briant KC, Allen A, Lin DW, Urban N, Drescher CW, Knudsen BS, Stirewalt DL, Gentleman R, Vessella RL, Nelson PS, Martin DB, Tewari M 2008 Circulating microRNAs as stable blood-based markers for cancer detection. *Proceedings of the National Academy of Sciences* **105**:10513-10518.